

# Fluorescence In Situ Hybridization (FISH)

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# Outline



- What is FISH.
- Principle of FISH
- Materials Required for FISH
- Types of probes
- Procedure of FISH
- Expected Results
- Safety Precautions

# ■ Objectives

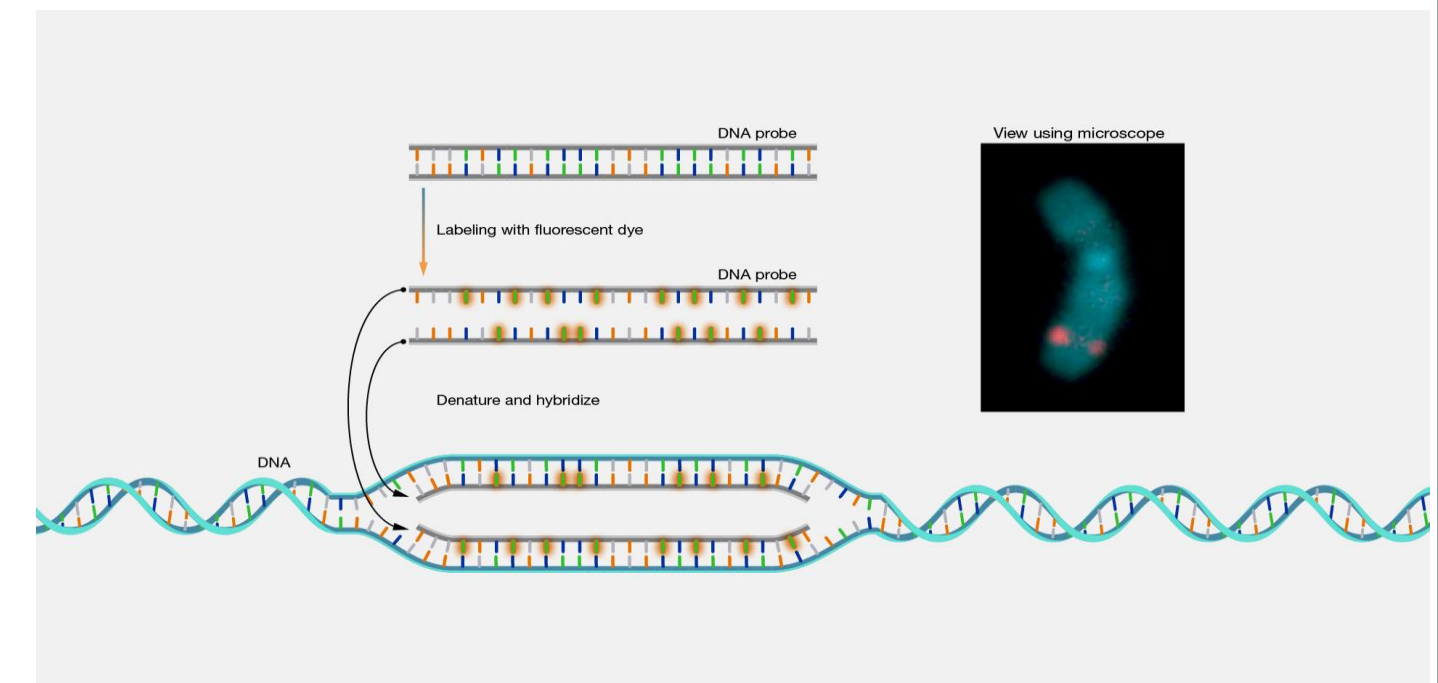


To acquaint students with the concepts of Fluorescence In Situ Hybridization.

# ❖ What is FISH



- Is a **laboratory technique** for **detecting and locating specific DNA sequences** on **chromosomes**.
- It is widely used in **research and clinical diagnostics** to identify **genetic abnormalities**, such as **chromosomal rearrangements**, **deletions**, **duplications**, and **aneuploidies**.



# ❖ Principle of FISH



- FISH is a technique that involves the hybridization of a fluorescently labelled **DNA or RNA probe** to its complementary target sequence within a biological sample.
- The hybridized probes are subsequently visualized using fluorescence microscopy, which enables the identification of **genetic abnormalities**, **gene mapping**, and **pathogen detection**.

## ❖ **Materials Required:**



1. **Sample:** Metaphase chromosomes from cells (blood, tissue, or cultured cells).
2. **Probe:** Fluorescently labeled DNA or RNA probe specific to the target sequence.
3. **Reagents:** Formamide or NaOH, Hybridization buffer, Wash buffers, Counterstain
4. **Equipment:** Fluorescence microscope, Water bath, Incubator and Slides and coverslips.

## ❖ Types of probes:

a



Gene-specific probe

b



Centromeric probe

c



Telomeric probe

d



Chromosome-painting probe

Repetitive-sequence probes

Examples of different types of fluorescence in situ hybridisation (FISH) probes

## ❖ Procedure of FISH:



1. **Sample Preparation:** Prepare metaphase chromosomes.
2. **Denaturation:** Denaturation solution (e.g., 70% formamide) at 70–75°C for 2–5 minutes, Dehydrate the slide using an ethanol series (70%, 85%, 100%) and air-dry.
3. **Probe Denaturation:** By heating it at 75°C for 5 minutes, then place it on ice.
4. **Hybridization:** Apply the denatured probe, incubate the slide in a humid chamber at 37°C for 4–16 hrs.

## **5. Post-Hybridization Washes:**

- Wash in formamide at 72°C for 2–5 minutes and Wash in formamide at room temperature for 1–2 minutes.

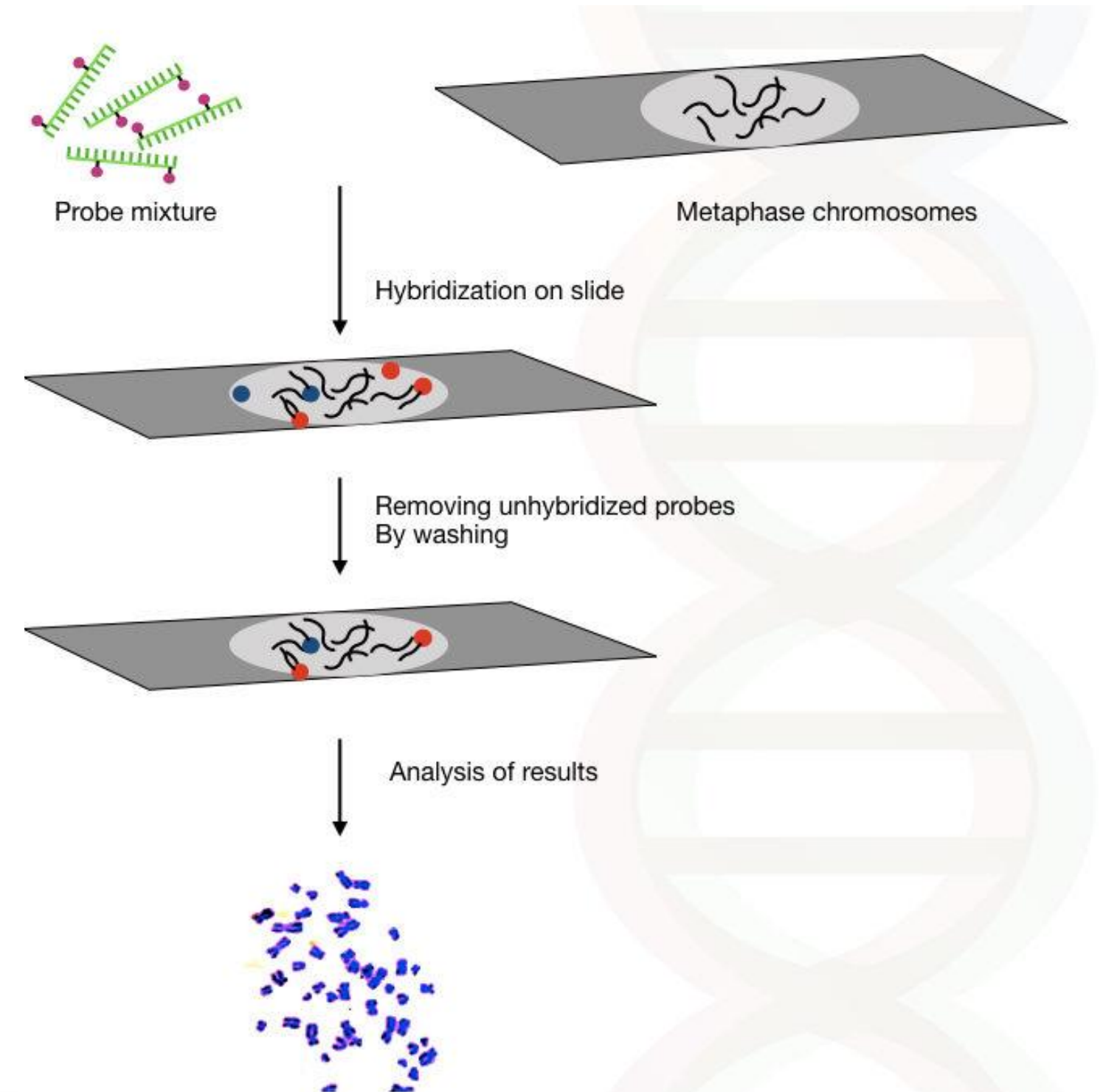
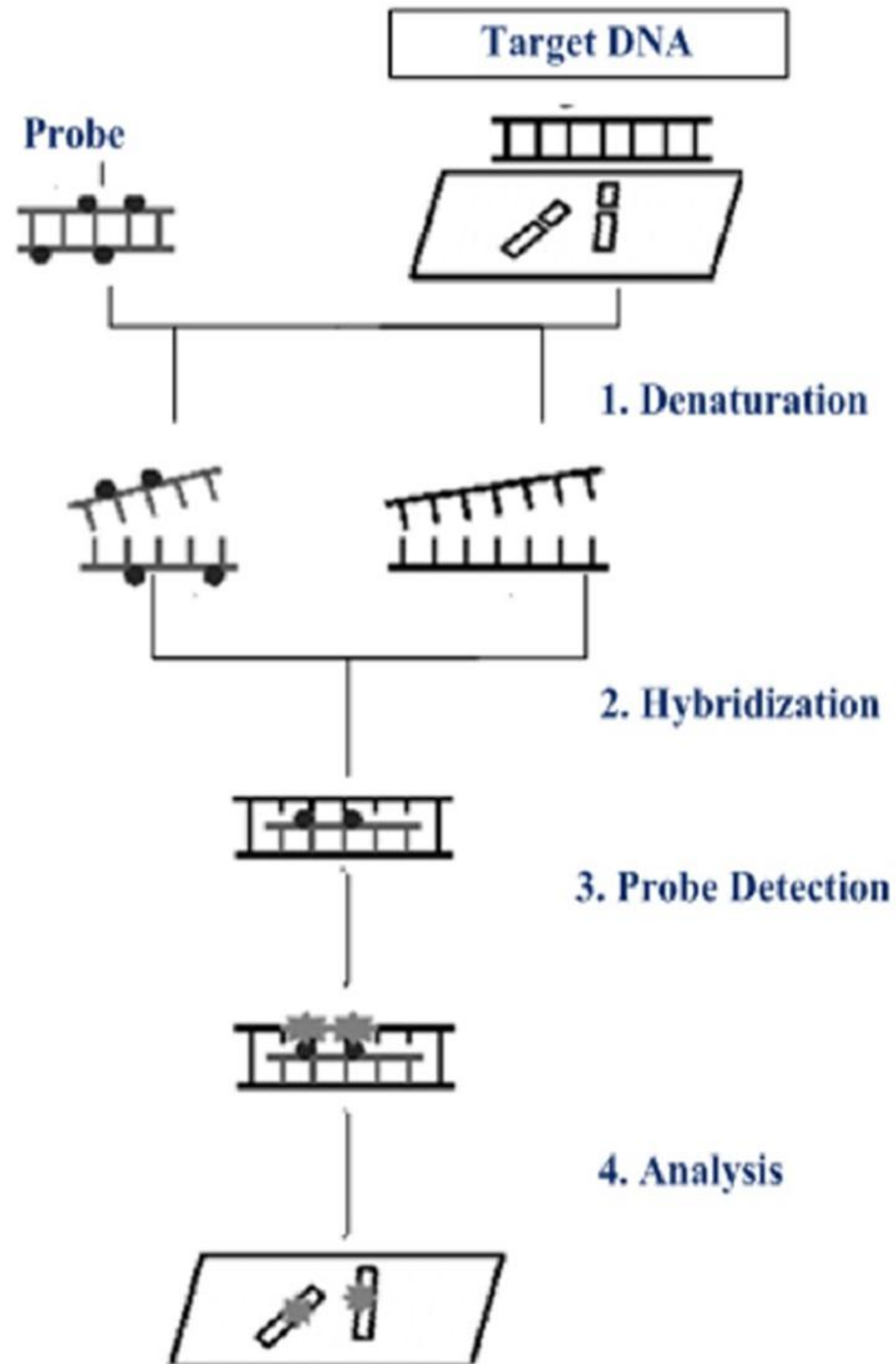
## **6. Counterstaining: .**

- Apply a counterstain, Mount the slide.

## **7. Visualization:**

- Observe the slide under a fluorescence microscope

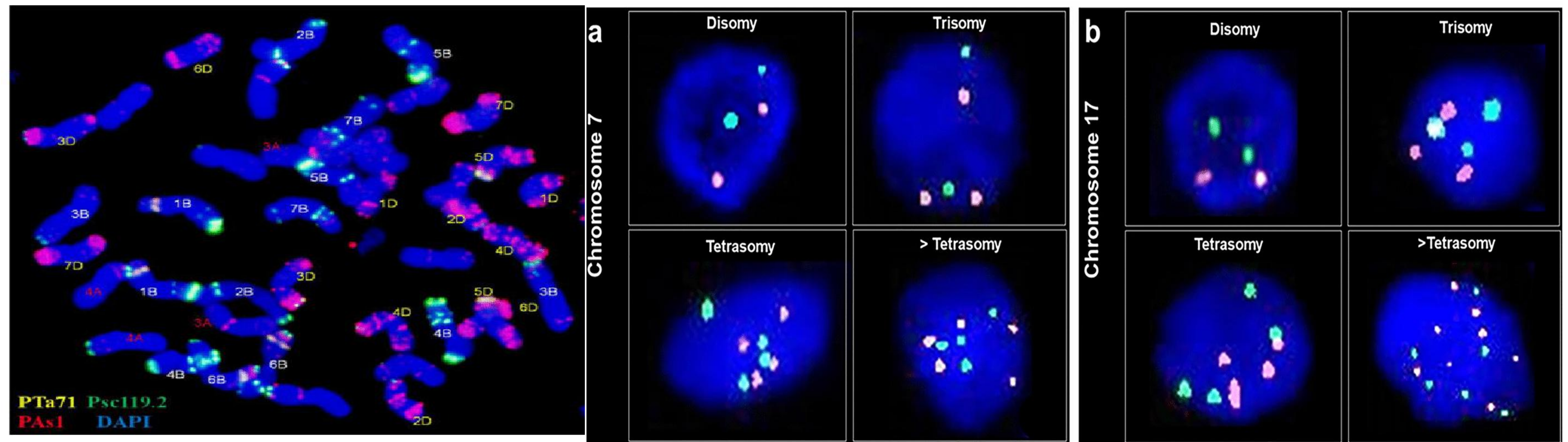
# Procedure of FISH:



# ❖ Expected Results:

## 1. Positive Signal:

- A fluorescent signal at the specific chromosomal location indicates the presence of the target sequence.

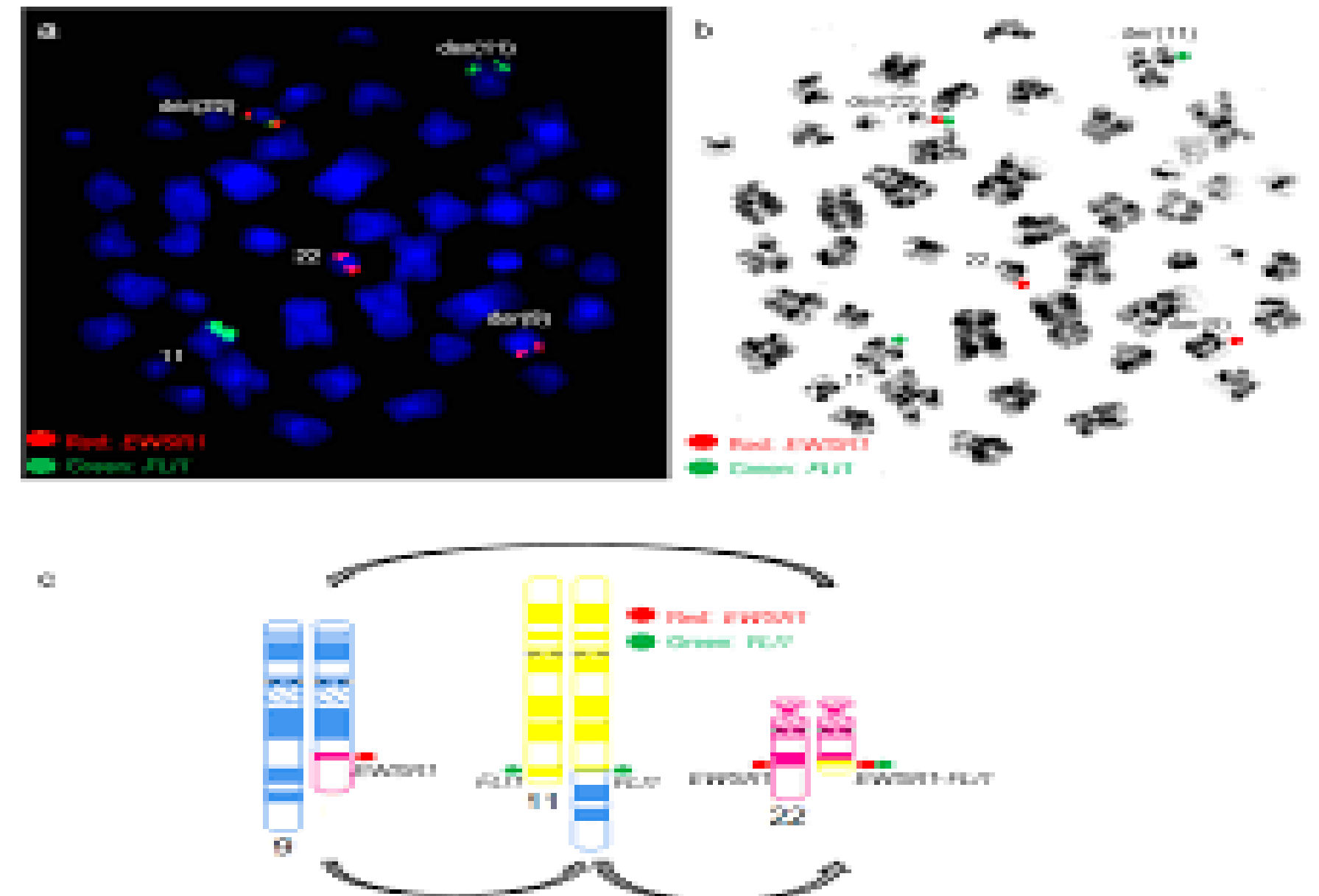


# ❖ Expected Results:



## 2. Negative Signal:

- Absence of a fluorescent signal indicates the absence of the target sequence.



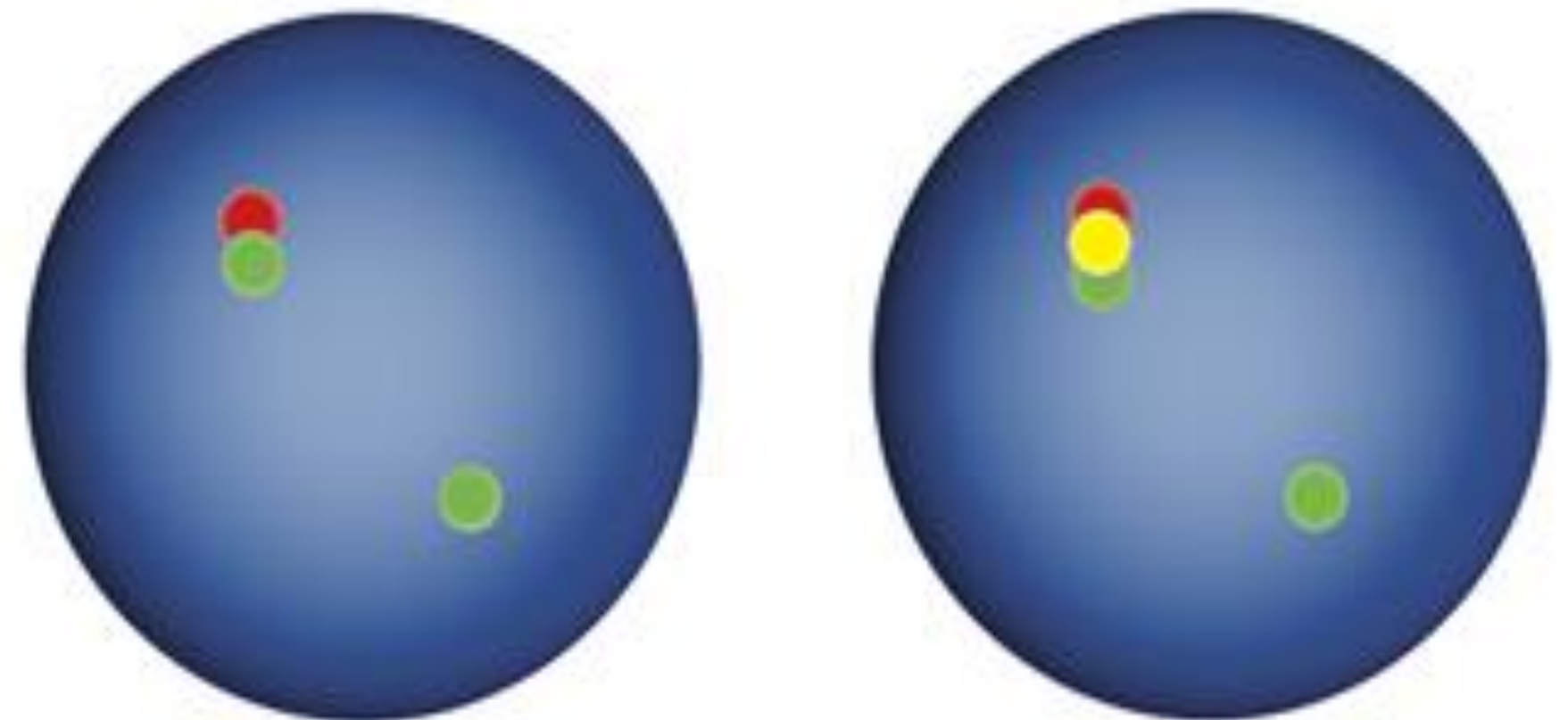
## ❖ Expected Results:



### 3. Abnormal Patterns:

- Abnormal patterns, such as split signals or additional signals, indicate chromosomal rearrangements or amplifications.

Abnormal



# ❖ Applications of FISH:



1. Cancer Diagnostics.
2. Prenatal Diagnosis
3. Genetic Research
4. Microbial Detection

## ❖ Advantages:



1. High specificity and sensitivity.
2. Can be used on non-dividing cells
3. Multiplexing capability

## ❖ Limitations:



1. Requires specialized equipment.
2. Time-consuming and labor-intensive
3. Probe design must be highly specific

## ❖ Safety Precautions:



- Handle fluorescence dyes with care as they may be hazardous.
- Use appropriate personal protective equipment .
- Dispose of reagents properly according to laboratory regulations.

# References



- Babcock, E. B., & Collins, J. L. (1918). Genetics laboratory manual. McGraw-Hill Book Company Incorporated.
- Blair, C. (2018). BIO2450L Genetics Laboratory Manual.
- VENNISON, S. (2009). Laboratory manual for genetic engineering. PHI Learning Pvt. Ltd..



**Thanks**